





#### Developing an individual-based model to study the bacterial denitrification process

Thesis submitted by

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for the degree of DOCTOR OF PHILOSOPHY by the Universitat Politècnica de Catalunya

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#### Context



- Abstraction of reality
- Simplification somehow
- Represent main features of reality



- Abstraction of reality
- Simplification somehow
- Represent main features of reality





- Abstraction of reality
- Simplification somehow
- Represent main features of reality
- Representation of a phenomenon
- Description of complex processes



Model

Reality

- Abstraction of reality
- Simplification somehow
- Represent main features of reality
- Representation of a phenomenon
- Description of complex processes
- Controllable environment
- Science lab







#### What is an Individual-Based Model?



### **Microbial IBMs**

- ✓ BACSIM (Kreft et al., 1998)
- ✓ INDISIM (Ginovart et al., 2002)
- ✓ MIOR (Masse et al., 2007)
- ✓ MICRODIMS (Verhulst et al., 2011)
- $\checkmark$  ..... Among others

#### **INDISIM: our core model**



# **Computational framework**

#### NETLOGO

- Open
- Widespread
- IBM
- Friendly use





### Denitrification







### **General Objective**

To develop an IBM to study the denitrification process driven by denitrifying bacteria, using thermodynamic principles to write microbial metabolic reactions as the centre of the individual sub-model. MbT-Tool: An open-access tool based on Thermodynamic Electron Equivalents Model to obtain microbialmetabolic reactions to be used in biotechnological process

# **Objectives**

To develop an open access and open source computational tool to systematize the writing of microbial metabolic reactions based on the thermodynamic principles to be used as the starting point of modelling projects dealing with biotechnological process carried out by microbes.

### What do we need?



### Thermodynamic approaches



### Thermodynamic approach



#### Thermodynamic approach (McCarty, 2007)



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## TEEM: Definitions (McCarty, 2007)

$$A = -\frac{\Delta G_s}{\varepsilon \Delta G_e} = \frac{\frac{(\Delta G_{in} - \Delta G_d)}{\varepsilon^n} + \frac{\Delta G_{pc}}{\varepsilon}}{\varepsilon(\Delta G_a - \Delta G_d)} = \frac{f_e^o}{f_s^o}$$

$$f_{s}^{o} = \frac{1}{1+A}$$
;  $f_{e}^{o} = \frac{A}{1+A}$ ;  $f_{s}^{o} + f_{e}^{o} = 1$ 

**FEEM 2**  

$$A = -\frac{\Delta G_s}{\varepsilon \Delta G_e} = \frac{\frac{(\Delta G_{fa} - \Delta G_d)}{\varepsilon^m} + \frac{(\Delta G_{in} - \Delta G_{fa})}{\varepsilon^n} + \frac{\Delta G_{pc}}{\varepsilon}}{\varepsilon \left(\Delta G_a - \Delta G_d - \frac{q}{p} \Delta G_{xy}\right)} = \frac{f_e^o}{f_s^o}$$

**TEEM 1** 

# **Energy-Transfer-Efficiency (**ε)

- Electrons distribution If  $\Delta G_{ic} < 1$ , then n = -1, and
  - Catabolism
  - Anabolism

$$\varepsilon = \left(\frac{\Delta G_{\rm pc}}{\Delta G_{\rm r} \left[1 - (\gamma_{\rm d}) / (\gamma_{\rm x} Y_{\rm C/C})\right] - \Delta G_{\rm ic}}\right)^{0.5}$$

- $\varepsilon = 0.37$
- Standard deviation error of 15%

McCarty, 2007

If  $\Delta G_{\rm ic} > 1$ , then n = +1, and  $\varepsilon = \left(\frac{\Delta G_{\rm pc} + \Delta G_{\rm ic}}{\Delta G_{\rm r} [1 - (\gamma_{\rm d})/(\gamma_{\rm x} Y_{\rm C/C})]}\right)^{0.5}$ 

#### Thermodynamic approach (McCarty, 2007)



### Software development





#### Software development





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#### MbT-Tool: Metabolism-based on Thermodynamics



#### MbT-Tool: Metabolism-based on Thermodynamics

Electron_donor		Microorganism	Microorganium	N-Sour
Glucose	7	C6.33H10.2103.53N - Saccharomyce	C6.33H10.2103.53N - Saccharomyces cerevisiae, glucose 🖤	NH4+
Chicose  Organic Reactions Acetate Alanine Benzoate Citrate Ethanol Formate Glucose Glutamate Glycerol Glycine Lactate Methane Methanol Palmitate Propionate Pyruvate Succinate NTA - Acid nitrilotriacetic Inorganic Reactions NH4+ -> NO2- NH4+ -> NO2- NH4+ -> N2 Fe2+ -> Fe3+ H2 -> H+ NO2> NO3- NO -> NO3-	v	S 33H10 2103 33N - Saccharomyce         N20 -> N0         N2 -> N20         H25 + H5> (S03)2-         H25 + H5> (S03)2-         (S03)2> (S04)2-         (S03)2> (S04)2-         (S203)2> (S04)2-         (S203)2> (S04)2-         H20 -> 02         NH4+         NH4+         NO3 -         NO2 -         N2	C5 31H10.2103.53h - Saccharomyces cerevisiae, glucose C5H702N - Casein, aerobic C7H1204N - Acetate, Ammomia N source aerobic C9H1505N - Acetate, Nitrate N source aerobic C9H1605N - Acetate, Nitrite N source aerobic C4.9H9.402.9N - Acetate, Methanogenic C4.9H903N - Glycine, Methanogenic C4.9H903N - Glycine, Methanogenic C4.9H903N - Glycine, Methanogenic C5H8.803.2N - Leucine, M C4.1H6.802.2N - Nutrient C5H8.803.2N - Leucine, M C4.1H6.802.2N - Nutrient C5H8.300.81N - Bacteria C5H8.3300.81N - Bacteria C5H8.20N - Bacteria, Unde C4.16H801.25N - Escherichia coli, undefined C3.85H6.6901.78N - Escherichia coli, glucose C6.33H10.2103.53N - Saccharomyces cerevisiae, glucose	NH4+
N2O -> NO3- NO + N2O -> NO3- N2 -> NO3- NO -> NO2- N2 -> NO2-			C4H7.2O1.93N - Paracoccus denitrificans, succinate C5H9O2.5N - Agrobacterium tumefaciens, succinate C4.17H8O1.75N - Bacteria, Undefined C(n)H(a)O(b)N(c) - Bacteria, Generic	

#### **MbT-Tool**

```
(rd) Electron donor --> NTA - Acid nitrilotriacetic :
 + 0.0556 NH4+ + 0.3333 HC03- + 1.1111 H+ + 1 e- --> + 0.0556 (C6H606N)3- + 0.6667 H20
 [\Delta G = 68.889 \text{ KJ/e-eq}]
(ra) Electron acceptor --> NO3- -> N2 :
 + 0.2 NO3- + 1.2 H+ + 1 e- --> + 0.1 N2 + 0.6 H20 [ \Delta G = -72.2 \text{ KJ/e-eq} ]
(rc) Biomass half reaction : C5H702N - Casein, aerobic , N-Source : NH4+
0.2 CO2 + 0.05 HCO3- + 0.05 NH4+ + 1 H+ + 1 e- --> 0.05 C'5'H'7'0'2'N'1 + 0.45 H2O
[\Delta G = 18.799 \text{ KJ/e-eq}]
                           _____
Energy reaction :
+ 0.0556 (C6H606N)3- + 0.0667 H20 + 0.2 N03- + 0.0889 H+ --> + 0.0556 NH4+ + 0.3333
HC03- + 0.1 N2
Synthesis reaction :
+ 0.0556 (C6H606N)3- + 0.2 C02 + 0.2167 H20 --> 0.05 C'5'H'7'0'2'N'1 + 0.1111 H+ +
0.0056 NH4+ + 0.2833 HC03-
Balanced equation using TEEM_1 :
[fe = 0.5] [fs = 0.5] [e = 0.33]
+ 0.0556 (C6H606N)3- + 0.1008 C02 + 0.1423 H20 + 0.0992 N03- --> 0.0252
C'5'H'7'0'2'N'1 + 0.0119 H+ + 0.0304 NH4+ + 0.3081 HC03- + 0.0496 N2
Yield prediction :
Yg/m = 51.311 [ grams_cells/mol_donor ]
Yc/m = 2.268 [ mol_C_cells/mol_donor ]
                                                                                 28
Yc/c = 0.378 [ mol_C_cell/mol_C_donor ]
    _____
```



Communication

#### MbT-Tool: An open-access tool based on Thermodynamic Electron Equivalents Model to obtain microbial-metabolic reactions to be used in biotechnological process

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INDISIM-Paracoccus: An individualbased and thermodynamic model to deal with *Paracoccus denitrificans* in a bioreactor

To use microbial metabolic reactions in the design, implementation and parameterization of the thermodynamic behaviour-rules embedded in the metabolic sub-model of an IBM for denitrifying bacteria in the framework of INDISIM, implementing the model in an open-access programming platform to achieve a simulator that facilitates exploring the effects of denitrifying bacterial metabolic sub-model.



\*(Xavier Portell, 2014) PhD Thesis presentation. Individual-based observations and individual-based simulations to study *Saccharomyces cerevisiae* cultures.



#### **INDISIM-Paracoccus submodels**



#### Cellular maintenance P. denitrificans





(Araujo et al., 2015) Thermodynamic Behavior-Rules for a Bacterial Individual-Based Model to Study the Denitrification Process (MathMod - February 2015 – IFAC, 2015- V.8 - pp: 743-748)
### **Microbial Metabolic Reactions P. denitrificans**

#	Microbial metabolic reactions (R)	ε
R1	$(C_4H_4O_4)^{2^2} + 0.60 \text{ NH}_4^* + 1.04 \text{ O}_2$ $\rightarrow 0.81 C_3H_{5.4}O_{1.45}N_{0.75} + 0.19 CO_2 + 1.40 \text{ HCO}_3^* + 0.34 \text{ H}_2\text{O}$	0.84
R2	$(C_4H_4O_4)^{2^-}$ + 0.08 NH <sub>4</sub> <sup>*</sup> + 0.52 NO <sub>3</sub> <sup>-</sup> + 1.05 H <sup>*</sup> + 0.18 H <sub>2</sub> O $\rightarrow$ 0.80 C <sub>3</sub> H <sub>5.4</sub> O <sub>1.45</sub> N <sub>0.75</sub> + 0.20 CO <sub>2</sub> + 1.40 HCO <sub>3</sub> <sup>-</sup>	0.90
R3	$(C_4H_4O_4)^{2^{\circ}} + 0.30 \text{ NH}_4^+ + 4.55 \text{ NO}_3^-)$ $\rightarrow 0.40 \text{ C}_3H_{5.4}O_{1.45}N_{0.75} + 4.55 \text{ NO}_2^- + 1.10 \text{ CO}_2 + 1.70 \text{ HCO}_3^- + 0.67 \text{ H}_2O$	0.41
R4	$(C_4H_4O_4)^{2^{\circ}}$ + 0.58 NH <sub>4</sub> <sup>*</sup> + 4.55 NO <sub>2</sub> <sup>*</sup> + 4.55 H <sup>*</sup> $\rightarrow$ 0.77 C <sub>3</sub> H <sub>5.4</sub> O <sub>1.45</sub> N <sub>0.75</sub> + 4.55 NO + 0.26 CO <sub>2</sub> + 1.42 HCO <sub>3</sub> <sup>*</sup> + 2.64 H <sub>2</sub> O	0.84
R5	$(C_4H_4O_4)^{2^{\circ}}$ + 0.58 NH <sub>4</sub> <sup>*</sup> + 4.55 NO $\rightarrow 0.77 C_3H_{5.4}O_{1.45}N_{0.75}$ + 2.28 N <sub>2</sub> O + 0.26 CO <sub>2</sub> + 1.42 HCO <sub>3</sub> <sup>*</sup> + 0.36 H <sub>2</sub> O	0.56
R6	$(C_4H_4O_4)^{2^{\circ}}$ + 0.58 NH <sub>4</sub> <sup>+</sup> + 2.28 N <sub>2</sub> O $\rightarrow 0.77 C_3H_{5.4}O_{1.45}N_{0.75}$ + 2.28 N <sub>2</sub> + 0.26 CO <sub>2</sub> + 1.42 HCO <sub>3</sub> <sup>-</sup> + 0.36 H <sub>2</sub> O	0.53

## Flow Diagram





## **Experimental conditions**



(Felgate et al., 2012) The impact of copper, nitrate and carbon status on the emission of nitrous oxide by two species of bacteria with biochemically distinct denitrification pathways.



(Felgate et al., 2012) The impact of copper, nitrate and carbon status on the emission of nitrous oxide by two species of bacteria with biochemically distinct denitrification pathways.

## **NetLogo implementation**



Implement the model

	Culture medium	Availability	Uptake-rate - u, (mol <sub>nutrient</sub> ·mol <sub>nass</sub> -1-h-1)		
Nutrient	concentroon	a, (h <sup>.1</sup> ) fixed	Tes	ting values	2
	al. (2022)	Dab	Low (L)	Medium (M)	High (H)
Succinate	5 20	0.28	0.065	0.13	0.52
Ammonium	[ල්]	Mal		-	0.31
Oxygen	236				0.54
Nitrate-a (aerobic)	- 983	Dodle	0.034	0.068	0.27
Nitrate-x (anaerobic)	@1.6095		0.019	0.119	1.19
Nitrite	255 - 0.0112	0.79	0.0062	0.062	0.62
Nitric Oxide		1.00	0.0000062	0.00062	0.62
Nitrous Oxide	0.003 - 0.000028	0.50	0.0031	0.031	0.31

Other bacterial parameters				
Parameter	Testing range	Calibrated value	Reference	
Cellular maintenance (gC <sub>donor</sub> ·gC <sub>mic</sub> ·1·h·1)		0.0020 - 0.0040	Gras et al. (2011) and van Verseveld et al. (1983)	
Mass split		0.50 (15% coefficient of variation)	Derived from (Ginovart et al., 2002a)	
Small bacterium size (µm)	0.4 - 0.6	0.5	Hall at all (1004)	
Big bacterium size (µm)	0.8 - 1.0	0.9	Holt et al. (1994)	
Minimum bacterium size al reproduc Biologic	cal pub	lished valu	Derived from Sas et al., 2011) and (Ginovart et al., 2002a)	



	Culture medium initial	Availability coefficient -	Uptake-rate - U/ (mol <sub>nutrient</sub> ·mol <sub>mass</sub> -1-h-1)			
Nutrient	concentration	a, (h <sup>-1</sup> ) fixed	Testing values			
	al. (2012)	Dab	Low (L)	Medium (M)	High (H)	
Succinate	5 - 20	0.28	0.065	0.13	0.52	
Ammonium	10	0.84		AND	S	
Oxygen	0.236	0.79		OF VELLO	0.54	
Nitrate-a (aerobic) Nitrate-x	4.9983 - 21.6095	0.63	TCSEU 0.019	0.068	0.27 1.19	
Nitrite	0.0255 - 0.0112	0.79	0.0062	0.062	0.62	
Nitric Oxide		1.00	0.0000062	0.00062	0.62	
Nitrous Oxide	0.003 - 0.000028	0.50	0.0031	0.031	0.31	

## Sensitivity Analysis – Aerobic phase



# **Calibration** Sensitivity Analysis – Anoxic phase



### e<sup>-</sup>-donor limited / e<sup>-</sup>-acceptor sufficient

# Calibration Sensitivity Analysis – Anoxic phase



### e<sup>-</sup>-donor sufficient / e<sup>-</sup>-acceptor limited 47

To use the simulator obtained to test hypotheses and diverse metabolic strategies for the individual behaviour of Paracoccus denitrificans in relation to the use of substrates growing in aerobic and anaerobic conditions in a bioreactor, testing the adequacy of the simulation outputs with experimental published data

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# Two hypotheses to test

The **bacterium prioritizes the use** of those nitrogen oxides with a higher degree of oxidation over others



# How did we evaluate our model?

- Factors: Specific Uptake-rate (each nutrient 6)
  - Aerobic phase: Succinate and NO<sub>3<sup>-</sup>(a)</sub>
  - Anaerobic phase: Succinate, NO<sub>3<sup>-</sup>(x)</sub>, NO<sub>2<sup>-</sup></sub>, NO and N<sub>2</sub>O
- **Levels**: 3
- Replicate: 3
- Response: Score
  - 7 time evolutions (2 aerobic and 5 anaerobic phases)
  - 2 Hypotheses



## **Rating the parameters combination**







### e<sup>-</sup>-donor limited / e<sup>-</sup>-acceptor sufficient



### e<sup>-</sup>-donor sufficient / e<sup>-</sup>-acceptor limited



	Culture medium	Availability coefficient -	Uptake-rate - U/ (mol <sub>nutrient</sub> ·mol <sub>mass</sub> -1·h-1)			
Nutrient	concentration [mM] Felgate et al. (2012)	a, (h <sup>-1</sup> ) fixed according to Dab	Testing values			Calibrated
			Low (L)	Medium (M)	High (H)	Values
Succinate	5 - 20	0.28	0.065	0.13	0.52	0.52
Ammonium	10	0.84			0.31	0.31
Oxygen	0.236	0.79			0.54	0.54
Nitrate-a (aerobic)	4.9983 -	0.63	0.034	0.068	0.27	0.27
Nitrate-x (anaerobic)	21.6095	1000	0.019	0.119	1.19	0.119
Nitrite	0.0255 - 0.0112	0.79	0.0062	0.062	0.62	0.062 - 0.62
Nitric Oxide		1.00	0.0000062	0.00062	0.62	0.62
Nitrous Oxide	0.003 - 0.000028	0.50	0.0031	0.031	0.31	0.31

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#### INDISIM-Paracoccus, an individual-based and thermodynamic model for a denitrifying bacterium



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#### HIGHLIGHTS

- An IBM to study denitrification that uses thermodynamics for the cellular activity.
- The simulator facilitates interaction between modelers and experts in denitrification.
- The thermodynamic properties embedded into individual cells for modeling.

#### G R A P H I C A L A B S T R A C T

The individual-based model approach with the thermodynamics embedded as an intracellular model defines the behavior-rule of the individual cell for maintenance and biomass generation to study the denitrification products dynamics, especially the greenhouse gas N<sub>2</sub>O, carried out by denitrifying bacterium Paracoccus denitrificans.



### http://dx.doi.org/10.1016/j.jtbi.2016.05.017

INDISIM-Denitrification: an individual-based model for study the denitrification process

# Objective

According to the results previously obtained, to improve the model design, modifying the individual rules required in the individual-based modelling context and to generalize the model to tackle other **denitrifying bacteria** using a wider set of published experimental data, performing the sensitivity analysis for some models' parameters in order to learn how the system works, and complete the modelling cycle.



### **INDISIM-Denitrification submodels**



## More ideas to extend the model

- **To include all thermodynamics calculations** into the implementation of the model
- To change the microbial biomass
  - Any denitrifying bacteria
- To test the model with two denitrifying bacteria with two experimental conditions and two bioreactor protocols
  - Paracoccus denitrificans
  - Achromobacter xylosoxidans

# Flow Diagram



## **NetLogo implementation**



## Metabolic pathways A. xylosoxidans

#	Microbial metabolic reactions (R)	3
R1	$(C_4H_4O_4)^{2^{-}} + 0.50 \text{ NH}_4^+ + 0.89 \text{ O}_2$ $\rightarrow 0.50 \text{ C}_5H_9O_{2.5}\text{N} + 0.013 \text{ CO}_2 + 1.50 \text{ HCO}_3^- + 0.006 \text{ H}_2\text{O}_3$	0.76
R2	$(C_4H_4O_4)^{2^-} + 0.77 \text{ NO}_3^+ + 1.54 \text{ H}^+ + 0.52 \text{ H}_2O$ $\rightarrow 0.37 \text{ C}_5H_9O_{2.5}\text{N} + 0.51 \text{ CO}_2 + 1.63 \text{ HCO}_3^- + 0.40 \text{ NH}_4^+$	0.65
R3	$(C_4H_4O_4)^{2^{\circ}} + 0.24 \text{ NH}_4^* + 4.49 \text{ NO}_3^{\circ} \rightarrow 0.24 \text{ C}_5H_9O_{2.5}\text{N} + 4.49 \text{ NO}_2^* + 1.05 \text{ CO}_2 + 1.76 \text{ HCO}_3^\circ + 0.52 \text{ H}_2\text{O}_3^\circ$	0.41
R4	$(C_4H_4O_4)^{2^{-}}$ + 0.45 NH <sub>4</sub> <sup>+</sup> + 4.54 NO <sub>2</sub> <sup>-</sup> + 4.54 H <sup>+</sup> $\rightarrow 0.45 C_5H_9O_{2.5}N$ + 4.54 NO + 0.20 CO <sub>2</sub> + 1.55 HCO <sub>3</sub> <sup>-</sup> + 2.37 H <sub>2</sub> O	0.84
R5	$(C_4H_4O_4)^2 + 0.50 \text{ NH}_4^* + 3.53 \text{ NO}$ $\rightarrow 0.50 \text{ C}_5H_9O_{2.5}\text{N} + 1.77 \text{ N}_2\text{O} + 0.006 \text{ CO}_2 + 1.50 \text{ HCO}_3^* + 0.006 \text{ H}_2\text{O}$	0.66
R6	$(C_4H_4O_4)^{2^-}$ + 0.24 NH <sub>4</sub> <sup>*</sup> + 4.50 N <sub>2</sub> O $\rightarrow 0.24 C_5H_9O_{2.5}N$ + 4.50 N <sub>2</sub> + 1.05 CO <sub>2</sub> + 1.76 HCO <sub>3</sub> <sup>-</sup> + 0.52 H <sub>2</sub> O	0.27

### Microbial Metabolic Reactions for individual mass degradation

Bacteria	Cytotoxic gas	Microbial metabolic reaction (Rg)
rificans	NO	$C_3H_{5.4}O_{1.45}N_{0.75} + {}^{49}/_4 \text{ NO}$ $\rightarrow {}^{9}/_4 \text{ CO}_2 + {}^{3}/_4 \text{ HCO}_3 + {}^{49}/_8 \text{ N}_2\text{ O} + {}^{3}/_4 \text{ NH}_4 + {}^{33}/_{40} \text{ H}_2\text{ O}$
P. denit	N <sub>2</sub> O	$C_3H_{5.4}O_{1.45}N_{0.75} + {}^{49}/_8N_2O$ $\rightarrow {}^{9}/_4CO_2 + {}^{3}/_4HCO_3 + {}^{49}/_8N_2 + {}^{3}/_4NH_4 + {}^{33}/_{40}H_2O$
oxidans	NO	$C_5H_9O_{2.5}N + 21 \text{ NO} \rightarrow 4 \text{ CO}_2 + \text{HCO}_3^- + {}^{21}/_2 \text{ N}_2\text{O} + \text{NH}_4^+ + 2 \text{ H}_2\text{O}$
A. xylos	N <sub>2</sub> O	$C_5H_9O_{2.5}N + {}^{21}/_2N_2O \rightarrow 4CO_2 + HCO_3 + {}^{21}/_2N_2 + NH_4 + 2H_2O_3$

(Araujo et al., 2016) Mass degradation to reduce cytotoxic products as an individual behavior-rule embedded in a microbial model for the study of the denitrification process (BioMicroWorld2015 – Proceedings book)

### P. denitrificans: e<sup>-</sup>-donor limited / e<sup>-</sup>-acceptor sufficient



### P. denitrificans: e<sup>-</sup>-donor sufficient / e<sup>-</sup>-acceptor limited



### A. xylosoxidans: e<sup>-</sup>-donor limited / e<sup>-</sup>-acceptor sufficient



### A. xylosoxidans: e<sup>-</sup>-donor sufficient / e<sup>-</sup>-acceptor limited





## 2016 @ASM Conference The Individual Microbe: Single-cell Analysis and Agent-based Modeling

March, 18<sup>th</sup> 2016







The Individual Microbe: Single-

Cell Analysis and Agent-Based



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Research Topic

# INDISIM-Denitrification: an individual-based model for study the denitrification process

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## **Future perspectives**



- The model can be adapted to more complex systems, e.g. wastewater treatments, soil management, and composting processes, among others.
- INDISIM-Denitrification could be incorporated into INDISIM-SOM, extending this soil model to complement the soil nitrogen cycle to deal with a mixed microbial community.
- There are some experimental works, which make reference to the role played by some elements in the denitrification process such is copper and/or iron, because they are a co-factor in activating some denitrifying enzymes. Study of this relation through the modeling process will be of great interest. Using a model such as INDISIM-Denitrification could be the next step to progress in knowledge of denitrification.

- We believe that the use of an approach in the field of nonequilibrium thermodynamics to describe the microbial metabolism has shown successful results and this methodology could be extended to other modelling frameworks.
- The use of MbT-tool outputs could be assumed as a starting point to design the metabolic sub models in other INDISIM branches to improve the design and parametrization of the model.
- Virtual experiments can be developed with some specific environmental characteristics where the bacteria execute a metabolic pathway using some value of energytransfer-efficiency while in another environmental condition it executes the same pathway using a different ε value.

## Conclusions

- An open access and open source tool has been developed to write microbial metabolic reactions based on Thermodynamic Electron Equivalents Model.
- The individual-based model named INDISIM-Paracoccus has been developed, including metabolic reactions as the basis of the individual behaviour-rules for the cellular maintenance and biomass synthesis sub-model, to deal with Paracoccus denitrificans growing in a bioreactor working as a batch and/or continuous culture. It has been verified that the corresponding simulator implemented in NetLogo platform works in accordance with its conceptual design.
- To improve the first simulator INDISIM-Paracoccus, a new individual-based model named INDISIM-Denitrification has been produced, which includes the new individual rule to reduce cytotoxic products, nitric oxide and/or nitrous oxide, through the degradation of individual mass.







## Thank you

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